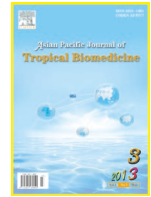




Contents lists available at ScienceDirect

Asian Pacific Journal of Tropical Biomedicine

journal homepage: www.elsevier.com/locate/apjtb

Document heading

doi:10.1016/S2221-1691(13)60059-3

© 2013 by the Asian Pacific Journal of Tropical Biomedicine. All rights reserved.

The development of *Terminalia chebula* Retz. (Combretaceae) in clinical research

Anwesa Bag, Subir Kumar Bhattacharyya, Rabi Ranjan Chattopadhyay*

Agricultural and Ecological Research Unit, Indian Statistical Institute 203, Barrackpore Trunk Road Kolkata-700 108, India

PEER REVIEW

Peer reviewer

Rumiza Abd Rashid, Chemistry (Forensic Analysis) Programme Faculty of Applied Sciences, Universiti Teknologi MARA 40450 Shah Alam, Selangor, Malaysia.

Tel: +603-55437855

Fax: +603-55444562

E-mail: rumiza9550@salam.uitm.edu.my

Comments

This paper is a good review paper on ayurvedic and pharmacological activities of *Terminalia chebula*. Citations used are also a good resources for reviewing and very informative to all the ayurvedic and traditional practitioners.

(Details on Page 250)

ABSTRACT

Medicinal plants are part and parcel of human society to combat diseases from the dawn of civilization. *Terminalia chebula* Retz. (Fam. Combretaceae), is called the 'King of Medicine' in Tibet and is always listed at the top of the list of 'Ayurvedic Materia Medica' because of its extraordinary power of healing. The whole plant possesses high medicinal value and traditionally used for the treatment of various ailments for human beings. Some of the folklore people used this plant in the treatment of asthma, sore throat, vomiting, hiccough, diarrhea, dysentery, bleeding piles, ulcers, gout, heart and bladder diseases. The plant has been demonstrated to possess multiple pharmacological and medicinal activities, such as antioxidant, antimicrobial, antidiabetic, hepatoprotective, anti-inflammatory, antimutagenic, antiproliferative, radioprotective, cardioprotective, antiarthritic, anticaries, gastrointestinal motility and wound healing activity. But no systematic updated information on the therapeutic effectiveness of *Terminalia chebula*, a popular herbal remedy in India and South-East Asia has so far been reported. This review highlights an updated information particularly on the phytochemistry and various pharmacological and medicinal properties of *Terminalia chebula* Retz. and some of its isolated compounds, along with their safety evaluation. This may provide incentive for proper evaluation of the plant as medicinal agent against the human diseases and also to bridge the lacunae in the existing literature and future scope which may offer immense opportunity for researchers engaged in validation of the traditional claims and development of safe and effective botanical medicine.

KEYWORDS

Terminalia chebula, Human diseases, Medicinal value, Bioactive constituents, Safety evaluation

1. Introduction

Ayurveda is a 5000 years old healing tradition rooted in ancient Indian culture. This vast body of healing knowledge –sometimes referred to as “Mother of all healing” – has recently come to the attention of Western medical researchers on seeking novel therapeutic compounds due to concerns over more invasive, expensive and potentially toxic main stream practices. According to World Health Organization, about 80% of world population rely chiefly on plant based traditional medicine for their primary healthcare need[1]. Traditional healing system around the world that

utilizes herbal remedies are an important resource for the discovery of modern drugs[2]. While screening a number of medicinal plants, scientist discovered one of the most revered medicinal plant *i.e.* *Terminalia chebula* (*T. chebula*) Retz. (Combretaceae), which exhibited a number of medicinal activities due to the presence of a large number of different types of phytoconstituents. The fruit of the tree possesses diverse health benefits and has been used as traditional medicine for household remedy against various human ailments since antiquity[3–5]. *T. chebula* has been extensively used in Ayurveda, Unani and Homoeopathic medicine and has become a cynosure of modern medicine.

*Corresponding author: Rabi Ranjan Chattopadhyay, Agricultural and Ecological Research Unit, Indian Statistical Institute 203, Barrackpore Trunk Road Kolkata-700 108, India.

Tel: +91-33-2575 3275

Fax: +91-33-2577 3049

E-mail: rabi@isical.ac.in; rabi.chattopadhyay@gmail.com

Foundation Project: Supported by Indian Statistical Institute, Kolkata, India (Grant No. Project A/C No. 5613).

Article history:

Received 15 Jan 2013

Received in revised form 27 Jan, 2nd revised form 5 Feb, 3rd revised form 12 Feb 2013

Accepted 26 Feb 2013

Available online 28 Mar 2013

The observed health benefits may be credited to the presence of the various phytochemicals like polyphenols, terpenes, anthocyanins, flavonoids, alkaloids and glycosides. The purpose of this review is to gather together the available published information on pharmacological and phytochemical analysis of the extracts and some of the isolated compounds of this plant as well as their toxic effects in a bid to highlighting the importance of this untapped resource in the fight against the human diseases.

2. *T. chebula* Retz.

2.1. Botanical description

The tree is tall about 50–80 feet in height. It has round crown and spreading branches. The bark is dark brown with some longitudinal cracks. Leaves are ovate and elliptical, with two large glands at the top of the petiole. The flowers are monoecious, dull white to yellow, with a strong unpleasant odour, borne in terminal spikes or short panicles. The flowers appear May–June, the fruits July–December. The fruit or drupe is about 1–2 inches in size. It has five lines or five ribs on the outer skin. Fruit is green when unripe and yellowish grey when ripe. Fruits were collected from January to April, fruit formation started from November to January^[6].

2.2. Special identity

Taxonomic description of *T. chebula* Retz. include Kingdom: Plantae–Plants; Subkingdom: Tracheobionta–Vascular plants; Superdivision: Spermatophyta–seed plants; Division: Magnoliophyta–flowering plants; Class: Magnoliopsida–dicotyledons; Subclass: Rosidae; Order: Myrtales; Family: Combretaceae–Indian almond family; Genus: Terminalia L–tropical almond; Species: *T. chebula* (Gaertn) Retz.–myrobalan.

Vernacular names of *T. chebula* Retz. include Assamese: shilikha; Bengali: haritaki; English: Chebulic myrobalan; Gujrati: hardi, harde; Hindi: hara; Kannada: alale; Konkani : ordo, hardi; Malayalam : katukka; Manipuri: Manali; Marathi: hirda; Oriya: karadha; Persian: halela; Sanskrit: haritaki; Sindhi: har; Tamil: Kata–K–Kay, Kadukkai; Telegu: Karaka; Urdu: Haejarad.

T. chebula is found in the Sub Himalayan tracks from Ravi eastwards to West Bengal and Assam, ascending upto the altitude of 1500 m in the Himalayas. This tree is wild in forests of Northern India, central provinces and Bengal, common in Madras, Mysore and in the southern part of the Bombay presidency^[7].

Classification according to size of the *T. chebula* fruit: Survari harade–which is large, dense, and heavy about 2 inches long, yellowish brown; Rangari harade–these is smaller, less wrinkled and less furrowed than the Survari

harade, in length about an inch; the epidermis is yellow; Bala harade–is smaller than the above two varieties, whose colour is deep brown to black; highly wrinkled, dark or brown epidermis; Java harade–these is the smallest of all, other characters are similar to those of Bala harade.

Classification according to the shape of the fruit: Vijaya–having alabu shape, used in all diseases, habitat in Vindhya mountain; Rohini–round in shape, used in vrana, habitat in Zansi and other parts of Madhya Pradesh; Pootana–size is small, mesocarp is less, seed is bigger, externally used, habitat Sindha; Amrita–Mesocarp is more used for shodhanakarma, habitat Madhya Pradesh and Champaranya; Abhya– fruit having 5 ribs, used in eye diseases, habitate Champaranya, Himalaya; Jeevantee–fruit is golden yellow, used in all diseases, habitate Himalaya; Chetaki–ruit having three ribs, used as purgative.

Classification according to the growth of the fruit: Zira –when the size is that of cumin seed; Javi–when the size is that of barley corn; Zangi–when the size is of a raisin; Chini–when the fruit is greenish yellow and somewhat hard; Asfer–when it is very nearly mature; Kabul–when it is fully matured^[8].

3. Ethnobotanical uses

The fruit is mild laxative, stomachic, tonic, alterative, antispasmodic. It is useful in ophthalmia, hemorrhoids, dental caries, bleeding gums, ulcerated oral cavity. Its paste with water is found to be anti-inflammatory, analgesic and having purifying and healing capacity for wounds. Its decoction is used as gargle in oral ulcers, sore throat. Its powder is a good astringent dentifrice in loose gums, bleeding and ulceration in gums. It is good to increase appetite, digestive aid, liver stimulant, stomachic, gastrointestinal prokinetic agent, and mild laxative. The powder of *T. chebula* fruits has been used in chronic diarrhea. It is used in nervous weakness, nervous irritability. It promotes the receiving power of five senses. It is adjuvant in hemorrhages due to its astringent nature and good for chronic cough, chorizo, sore throat as well as asthma. Also it is useful in renal calculi, dysurea, retention of urine and skin disorders with discharges like allergies, urticaria and other erythematous disorders^[4,9].

4. Diseases that have beneficial effects

Digestive diseases, urinary diseases, diabetes, skin diseases, heart diseases, irregular fevers, constipation, ulcers, vomiting, colic pain, haemorrhoids.

5. Phytoconstituents of *T. chebula* Retz.

The fruits of *T. chebula* is rich in tannins (about 32%–34%)

and its content varies with geographical distribution^[10,11]. The tannins of *T. chebula* are of pyrogallol (hydrolysable) type. A group of researchers found 14 components of hydrolysable tannins (gallic acid, chebulagic acid, punicalagin, chebulanin, corilagin, neochebulinic acid, ellagic acid, chebulinic acid, 1,2,3,4,6-penta-O-galloyl- β -D-glucose, 1,6-di-O-galloyl-D-glucose, casuarinin, 3,4,6-tri-O-galloyl-D-glucose, terchebulin) from *T. chebula* fruits^[12]. Other constituents include phenolics such as chebulinic acid, ellagic acid and anthraquinones. Some of the other minor constituents were polyphenols such as corilagin, galloyl glucose, punicalagin, terflavin A, maslinic acid^[13]. Besides, fructose, amino acids, succinic acid, betasitosterol, resin and purgative principle of anthraquinone are also present^[14,15]. Flavonol, glycosides, triterpenoids, coumarin conjugated with gallic acids called chebulin as well as other phenolic compounds were also isolated^[13,16–18]. Twelve fatty acids were isolated from *T. chebula* of which palmitic acid, linoleic acid and oleic acid were main constituents^[19]. Triterpenoid glycosides such as chebulosides I and II, arjunin, arjunglucoside, 2 α -hydroxyursolic acid and 2 α -hydroxymicromiric acid also have been reported^[20]. The leaves were found to contain polyphenols such as punicalin, punicalagin, terflavins B, C, and D^[12,21,22]. The plant is found to contain phloroglucimol and pyrogallol, along with phenolic acids such as ferulic, p-coumaric, caffeic and vanillic acids^[23]. Oil extracted from kernels yielded palmitic, stearic, oleic, linoleic, behenic and arachidic acids^[23].

6. Pharmacological activity

6.1. Antioxidant and free radical scavenging activity

The leaves, bark and fruit of *T. chebula* possessed high antioxidant activity and phenolics were found to be responsible for this activity^[24]. Aqueous extract of *T. chebula* inhibited xanthine/xanthine oxidase activity and was also an excellent scavenger of DPPH radicals^[25]. *T. chebula* in a polyherbal formulation (Aller-7/ NR-A2) inhibited free radical induced hemolysis and also significantly inhibited nitric oxide release from lipopolysaccharide stimulated murine macrophages^[26]. Six extracts and four compounds of *T. chebula* fruit exhibited antioxidant activity at different magnitudes of potency^[27]. Strong antioxidant activity of aqueous extract of *T. chebula* was observed by studying the inhibition of radiation induced lipid peroxidation in rat liver microsomes at different doses^[28], and methanolic extract was also found to inhibit lipid peroxide formation and to scavenge hydroxyl and superoxide radicals *in vitro*^[29]. Acetone extract has stronger antioxidant activity than α -tocopherol and HPLC analysis with diode array detection indicated the presence of hydroxybenzoic acid derivatives, hydroxycinnamic acid derivatives, flavonol aglycones and

their glycosides, as main phenolic compounds^[30].

6.2. Anticarcinogenic activity

A group of researchers have reported the inhibitory action on cancer cell growth by the phenolics of *T. chebula* Retz fruit and found that chebulinic acid, tannic acid and ellagic acid were the most growth inhibitory phenolics of *T. chebula*^[31]. Ethanol extract of *T. chebula* fruit inhibited cell proliferation and induced cell death in a dose dependent manner in several malignant cell lines including human (MCF-7) and mouse (S115) breast cancer cell line, human osteosarcoma cell line (HOS-1), human prostate cancer cell (PC-3) and a non-tumorigenic immortalized human prostate cell line (PNT1A)^[32]. Besides, acetone extract of bark and fruit powder of *T. chebula* harbors constituents with promising anticarcinogenic activity^[32]. Some pharmacological activities of *T. chebula* Retz. are shown in Table 1 and some isolated compounds from *T. chebula* Retz. with their bioactivities are shown in Table 2.

Table 1
Various pharmacological activities of *T. chebula* Retz.

Pharmacological activities	Reference(s)
Antioxidant	24–30
Antibacterial	47–58
Antifungal	59–62
Antiviral	63–70
Antiprotozoal	71–73
Anticarcinogenic	31,32
Radioprotective	25,34
Antimutagenic	33
Chemopreventive	35
Hepatoprotective	36–38
Cardioprotective	39,40
Cytoprotective	41–44
Antidiabetic	45, 46
Renoprotective	45
Antiinflammatory	74,76
Antiarthritic	75
Adaptogenic	77
Antianaphylactic	78
Hypolipidemic	79
Hypocholesterolemic	80
Gastrointestinal motility	81
Antiulcer	82
Antispasmodic	83
Anticaries	53,84
Wound healing	85
Purgative	86
Antiallergic	76
Immunomodulatory	87

6.3. Antimutagenic, radioprotective and chemopreventive activity

Antimutagenic activity of aqueous extract and hydrolyzable tannins from *T. chebula* in *Salmonella typhimurium* has been documented^[33]. Gamma radiation

Table 2Structure and activities of some active compounds and their derivatives from *T. chebula* Retz.

Class of compounds	Compounds	Plant parts	Activities	References
Phenolic acid	Ellagic acid	Fruit	Antibacterial activity against intestinal bacteria <i>Clostridium perfringes</i> , <i>Escherichia coli</i> ; antiproliferative activity	31,50
Phenolic acid	Chebulic acid	Fruit	Hepatoprotective, antioxidant and free radical scavenging activity, cytoprotective	36,41
Phenolic acid	Neochebulic acid	Fruit	Hepatoprotective, antioxidant and free radical scavenging activity	36
Phenolic acid	Gallic acid	Fruit	Antioxidant, antibacterial, antiviral, cytoprotective activity	41,51,63
Phenolics	2, 4–chebulyl–beta–D–glucopyranose	Fruit	Antiproliferative activity	31
Phenolic acid	Chebulinic acid	Fruit	Antiproliferative activity	31
Benzoic acid	Hydroxybenzoic acid derivatives	Fruit	Antioxidant activity	30
Cinnamic acid	Hydroxycinnamic acid derivatives	Fruit	Antioxidant activity	30
Flavonoid	Falvonol aglycones	Fruit	Antioxidant activity	30
Glycosides		Fruit	Antioxidant, antibacterial activity	30,50
Phenolic acid	Chebulagic acid	Fruit, Seed	Cytoprotective, anti–arthritic activity	41,75

induced strand breaks formation in plasmid PBR322 DNA was inhibited by aqueous extract of *T. chebula*[25]. The administration of aqueous extract of *T. chebula* prior to whole body irradiation of mice resulted in a reduction of peroxidation of membrane lipids in the mice liver as well as a decrease in radiation induced damage to DNA. It also protected the human lymphocytes from undergoing the gamma radiation–induced damage to DNA exposed *in vitro*[34]. *T. chebula* showed chemopreventive effect on nickel chloride –induced renal oxidative stress, toxicity and cell proliferation response in male Wistar rats[35].

6.4. Hepatoprotective activity

A mixture of chebulic acid (CA) and its minor isomer, neochebulic acid with a ratio of 2:1 isolated from ethanolic extract of *T. chebula* fruits showed strong hepatoprotective activity[36]. Ethanol extract *T. chebula* was found to prevent the hepatotoxicity caused by the administration of rifampicin, isoniazid and pyrazinamide (combination) in sub–chronic model (12 weeks)[37]. Protective effects of an aqueous extract of *T. chebula* fruit on the tert–butyl hydroperoxide–induced oxidative injury was observed in cultured rat primary hepatocytes and rat liver have also been documented[28,29]. *T. chebula* in a herbal formulation (HP–1) showed hepatoprotective activity against carbon tetrachloride induced toxicity in rat hepatocytes[38].

6.5. Cardioprotective activity

T. chebula extract pretreatment was found to ameliorate the effect of isoproterenol on lipid peroxide formation and retained the activities of the diagnostic marker enzymes in isoproterenol induced myocardial damage in rats[39]. Its pericap has also been reported to have cardioprotective activity in isolated frog heart model[40].

6.6. Cytoprotective activity

Gallic acid (GA) and CA were isolated from the extract of the herbal medicine Kashi (myrobalan, the fruit of *T. chebula*) as active principal that blocked the cytotoxic T–lymphocyte–mediated cytotoxicity. Granule exocytosis in response to anti–CD3 stimulation was also blocked by GA and CA at the equivalent concentrations[41]. The ethanolic extract of *T. chebula* fruit exhibited a notable cytoprotective effect on the HEK–N/F cells. In addition its extract exhibited significant cytoprotective effect against UV–induced oxidative damage. These observations were attributed to the inhibitory effect of the *T. chebula* extract on the age dependent shortening of the telomere length as shown by the Southern Blots of the terminal restriction fragments of DNA extracted from sub–culture passages[42]. It exhibited the development of duodenal ulcers and appeared to exert a cytoprotective effect on the gastric mucosa *in vitro*[43]. Cytoprotective effect on oxidative stress and inhibitory effect on cellular aging of its fruits have also been documented[44].

6.7. Antidiabetic and renoprotective activity

T. chebula fruit and seeds exhibited dose dependent reduction in blood glucose of streptozotocin induced diabetic rats both in short term and long term study and also had renoprotective activity[45,46].

6.8. Antibacterial activity

T. chebula exhibited antibacterial activity against a number of both Gram–positive and Gram–negative human pathogenic bacteria[47–49]. Ethanedioic acid and ellagic acid isolated from butanol fraction of *T. chebula* fruit extract had strong antibacterial activity against intestinal bacteria, *Clostridium perfringens* and *Escherichia coli*[50]. It is effective in inhibiting the urease activity of *Helicobacter pylori*,

an ubiquitous bacterium implicated in the development of gastritis, ulcers and stomach cancers[49]. GA and its ethyl ester isolated from ethanolic extract of *T. chebula* showed antimicrobial activity against methicillin-resistant *Staphylococcus aureus* (*S. aureus*) [51]. Ripe seeds of *T. chebula* also exhibited strong antibacterial activity against *S. aureus* [52]. The aqueous extract of *T. chebula* strongly inhibited the growth of *Streptococcus mutans*, salivary bacterial [53]. Diffusate of *T. chebula* showed an inhibitory effect against strain X-100 of the bacterium *Xanthomonas campestris* pv. citri indicating its usefulness for the management of citrus canker disease [54]. It has also growth inhibitory action against *Salmonella typhi* [55], *Klebsiella* [56], *Shigella* [47] and intestinal bacterial [50]. Ethanol extract of *T. chebula* fruit showed strong antibacterial activity against multidrug-resistant uropathogenic *Escherichia coli* and phenolics were found to be responsible for this antibacterial activity [57,58].

6.9. Antifungal activity

An aqueous extract of *T. chebula* exhibited antifungal activity against a number of dermatophytes and yeasts [59,60]. It is effective against the pathogenic yeast *Candida albicans* and dermatophytes *Epidermophyton*, *Floccosum*, *Microsporum gypseum* and *Trichophyton rubrum* [59]. Its inhibitory effect on three dermatophytes (*Trichophyton* spp.) and three yeasts (*Candida* spp.) has also been documented [61]. An aqueous extract of galls of *T. chebula* showed inhibitory effects on three dermatophytes (*Trichophyton* spp.) and three yeasts (*Candida* spp.) [59]. *In vitro* anticandidal activity of methanol extract of *T. chebula* was observed against clotrimazole resistant *Candida albicans* [62]. Seed extract exhibited antifungal activity against *Trichophyton glabrata* [59].

6.10. Antiviral activity

T. chebula fruits afforded four immunodeficiency virus type 1 (HIV-1) integrase inhibitors, GA (I) and three galloyl glucoses (II-IV). Their galloyl moiety plays a major role for inhibition against the 3'-processing of HIV-1 integrase of the compounds [63]. *T. chebula* has also retroviral reverse transcriptase inhibitory activity [64]. It protects epithelial cells against influenza A virus, supporting its traditional use for aiding in recovery from acute respiratory infections [65]. The methanol and aqueous extracts of *T. chebula* showed a significant inhibitory activity with $IC_{50} \leq 5 \mu\text{g/mL}$ on human immunodeficiency virus-1 reverse transcriptase [66]. It also demonstrated the therapeutic activity against herpes simplex virus both *in vitro* and *in vivo* tests [67]. These finding prompted a team of Japanese researchers to investigate *T. chebula's* effect on human cytomegalovirus (CMV). They found that *T. chebula* was effective in inhibiting the replication of human cytomegalovirus *in vitro* and in an

AIDS model with immunosuppressed mice and concluded that it may be beneficial for the prevention of CMV diseases and immunocompromised patients [68]. It is also helpful in sexually transmitted diseases and AIDS [69]. Tannins from *T. chebula* are effective against potato virus x [70].

6.11. Antiprotozoal activity

A combination of *T. chebula* and four other botanicals (*Boerhavia diffusa*, *Berberis aristata*, *Tinospora cordifolia*, and *Zingiber officinale*) had a maximum cure rate of 73% in experimental amoebic liver abscess in hamsters [71] and 89% in experimental caecal amoebiasis in rats showing its antiamoebic activity against *Entamoeba histolytica* [72]. The acetone extract of *T. chebula* seeds showed anti plasmodial activity against *Plasmodium falciparum* [73].

6.12. Anti-inflammatory and anti-arthritic activity

Aqueous extract of dried fruit of *T. chebula* showed anti-inflammatory by inhibiting inducible nitric oxide synthesis [74]. Chebulagic acid from immature seeds of *T. chebula* significantly suppressed the onset and progression of collagen induced arthritis in mice [75]. *T. chebula* in a polyherbal formulation (Aller-7) exhibited a dose dependent anti-inflammatory effect against Freund's adjuvant induced arthritis in rats [76].

6.13. Adaptogenic and antianaphylactic activities

T. chebula fruit was one of the six Ayurvedic herbs administered to animals to test their adaptogenic potential. All six traditional rasayana plants were able to aid the animals against a variety of different stressors working in different ways [77]. Besides, animal studies show that when extract of *T. chebula* was administered following induction of anaphylactic shock, the serum histamine levels were reduced, indicating its strong antianaphylactic action [78]. Water soluble fraction of *T. chebula* had a significant increasing effect on anti-dinitrophenyl IgE-induced tumor necrosis factor- α production from rat peritoneal mast cells indicating its strong antianaphylactic action [78].

6.14. Hypolipidemic and hypocholesterolemic activity

Hypolipidemic activity of *T. chebula* extract against experimentally induced atherosclerosis have been documented [79]. It also possessed hypocholesterolemic activity against cholesterol-induced hypercholesterolemia and atherosclerosis in rabbits [80].

6.15. Gastrointestinal motility improving and anti-ulcerogenic activity

Although its traditional use as laxative is well established,

T. chebula fruit has been shown to increase gastric emptying time^[81]. This action appeared to be balanced with a protective effect on the gastrointestinal mucosa, with the improvement in the secretory status of Brunner's gland involved in the protection against duodenal ulcer^[82].

6.16. Antispasmodic activity

One of the numerous studies of *T. chebula* demonstrated its 'anti-vata' or 'anti-spasmodic' properties by the reduction of abnormal blood pressure as well as intestinal spasms. This confirms its traditional usefulness for spastic colon and other intestinal disorders^[83].

6.17. Anticaries activity

The aqueous extract of *T. chebula* strongly inhibited the growth, sucrose induced adherence and glucan induced aggregation of *Streptococcus mutans*. Mouth rinsing with a 10% solution of the extract inhibited the salivary bacterial count and glycolysis of salivary bacteria for up to 90 min post rinsing^[53, 84].

6.18. Wound healing activity

Topical administration of an alcoholic extract of *T. chebula* leaves on the healing of rat dermal wounds showed that *T. chebula* treated wounds healed faster as indicated by improved rates of contraction and decreased period of epithelialization^[85].

6.19. Purgative property

Purgative action of an oil fraction from *T. chebula* has been documented^[86].

6.20. Immunomodulatory activity

Aqueous extract of *T. chebula* produced an increase in humoral antibody titer and delayed type hypersensitivity in mice^[87]. Crude extract of *T. chebula* stimulated cell-mediated immune response in experimental amoebic liver abscess in golden hamsters^[71].

6.21. Anti-allergic activity

Aller-7, a polyherbal formulation of seven medicinal plants including *T. chebula* exhibited potent *in vitro* antiallergic activity in isolated guinea pig ileum substrate^[76].

7. Clinical studies

Oral rinsing with extract of *T. chebula* was found to

significantly reduce both total bacterial counts and streptococcal counts in saliva samples. The protective effect lasted for about 3 h after rinsing, demonstrating a potential role of *T. chebula* in the prevention of dental caries^[53].

A short term clinical trial has been carried out on patients with simple constipation. *T. chebula* increases the stools and has got property of evacuating the bowel completely^[88].

Besides, some Ayurvedic drugs, consisting of *T. chebula* as one of the constituents have been subjected to clinical trials regarding their effects on constipation, mental and physical disability, allergic rhinitis and mental stress. In all the cases *T. chebula* containing drugs showed good effects in the treated groups when compared to their normal control patients^[89,90].

8. Safety evaluation

From the literature it has been noted that *T. chebula* exhibited significant hepatoprotective^[36–38], cardioprotective^[39,40], antimutagenic/anticarcinogenic^[31–33], cytoprotective^[41–44], antioxidant^[24–30] and adaptogenic^[77,78] effects. Aqueous, ethanol, and ethyl acetate extracts of *T. chebula* fruits also demonstrated no cellular toxicity on sheep erythrocytes as well as acute oral toxic effects on rats at recommended and higher doses^[48,91,92]. Besides, hydroalcoholic extract of *T. chebula* fruits demonstrated cytochrome P-450 inhibition potential in rats^[93]. *T. chebula* by itself had no genotoxic effect both in VITOTOX test and Ames assay^[94]. Rather, *T. chebula* fruit could reduce the lead and aluminium induced genotoxicity^[95,96]. The hydrolysable tannins obtained from *T. chebula* fruits also showed antimutagenic activity against direct acting mutagens like sodium azide and 4-nitro-O-phenylene diamine. These findings indicated that it is a safe substance to be used as drug ordinarily.

9. Conclusions and recommendations

T. chebula is one of the most versatile plants having a wide spectrum of pharmacological and medicinal activities. This versatile medicinal plant is the unique source of various types of compounds having diverse chemical structure. Though it has a number of pharmacological activities due to the presence of various types of bioactive compounds, very little work has been done on the plausible medicinal applications of this plant against the diseases particularly on multidrug resistant bacterial pathogens. Hence extensive investigation is needed to exploit their therapeutic ability to combat diseases including drug resistant infections. As the global scenario is now changing towards the use of nontoxic plant products having traditional medicinal use,

a drug development programme should be undertaken to develop modern drugs with the compounds isolated from *T. chebula* effective against different types of diseases and also to overcome the problem of drug resistance after extensive investigation of its bioactivity, mechanism of action, pharmacotherapeutics, toxicity and after proper standardization and clinical trials.

Conflict of interest statement

We declare that we have no conflict of interest.

Acknowledgements

Authors wish to acknowledge the Head, Agricultural and Ecological Research Unit, Indian Statistical Institute, Kolkata, India for her kind help and cooperation during this work and also Prof. T. K. Basu, former Head, Biometry Research Unit, Indian Statistical Institute, Kolkata, India for critically going through the manuscript.

Comments

Background

This is a review paper on the benefits of *T. chebula* as an alternative medicine for many diseases. The pharmacological effects exhibited by this plant have been elaborated in depth with citations from studies that have been conducted using this Ayurvedic plant.

Research frontiers

There is no lab experiment being done in this manuscript since it is a review paper. However, the author cited latest and recent publications on works done in this particular field, in which bring the readers to the recent analytical approach for pharmacological potential of this plant.

Related reports

The author cited different papers in his manuscript to support the therapeutic potential of *T. chebula* in traditional medicine. Past studies mostly presented the pharmacological activities of this plant done *in vitro* and *in vivo*.

Innovations and breakthroughs

This review paper is one of its own in which it summarizes any research that have been conducted on *T. chebula* specifically in medicinal field. It is a good source of literature survey for researchers who intended to do studies in this particular field, and using this plant.

Applications

This paper could be applied by most Ayurvedic practitioners in their medication activities to treat patients with different types of diseases.

Peer review

This paper is a good review paper on Ayurvedic and pharmacological activities of *T. chebula*. Citations used are also a good resources for reviewing and very informative to all the Ayurvedic and traditional practitioners.

References

- [1] World Health Organization. *Traditional medicine—growing needs and potential. WHO policy perspectives on medicine, No. 2. WHO/EBM/2002*. WHO: Geneva; 2002.
- [2] Koehn FE, Carter GT. The evolving role of natural products in drug discovery. *Nat Rev Drug Discov* 2005; **4**: 206–220.
- [3] CSIR. *The wealth of India – A dictionary of Indian raw materials and industrial products. Vol X*. New Delhi: Publication and Information Directorate, CSIR; 2002, p. 522–524.
- [4] Varier. *A dictionary of Indian raw materials and industrial products*. New Delhi: Publications and Information Directorate, Council of Scientific and Industrial Research; 2002, p. 387.
- [5] Khare CP. *Indian medicinal plants: An illustrated dictionary*. Berlin: Springer-Verlag; 2007, p. 652–653.
- [6] Govt. of India. *The Ayurvedic pharmacopoeia of India*. New Delhi: Government of India Ministry of Health and Family Welfare Department of Indian System of Medicine & Homoeopathy; 2001, p. 47, 143.
- [7] Gupta AK, Tandon N, Sharma M. *Quality standards of Indian medicinal plant*. New Delhi: Indian Council of Medical Research; 2003, p. 207–209.
- [8] Sukhdev SH, Deepak M, Joseph GVR, Joseph S, Nagar G. *Indian herbal pharmacopoeia. Vol II*. Jammu Tawi: IDM, Mumbai and RRL, CSIR; 1999, p. 154–159.
- [9] Aslokar LV, Kakkar KK, Chakre OJ. *Glossary of Indian medicinal plants with active principles*. New Delhi: Publications and Information's Directorate, CSIR; 1992.
- [10] Kumar A, Lakshman K, Jayaveera K, Satish K, Tripathi SM. Estimation of rutin and quercetin *Terminalia chebula* by HPLC. *Int J Aesth Antiag Med* 2009; **2**(1): 3.
- [11] Jayaramkumar K. Effect of geographical variation on content of tannic acid, gallic acid, chebulinic acid, and ethyl gallate in *Terminalia chebula* fruits. *Nat Prod* 2006; **2**(3–4): 170–175.
- [12] Juang LJ, Sheu SJ, Lin TC. Determination of hydrolyzable tannins in the fruit of *Terminalia chebula* by high-performance liquid chromatography and capillary electrophoresis. *J Sep Sci* 2004; **27**(9): 718–724.
- [13] Williamson EN. *Major herbs of Ayurveda*. London: Churchill Livingstone; 2002, p. 299.
- [14] Tubtimdee C, Shotipruk A. Extraction of phenolics from *Terminalia chebula* Retz. with water-ethanol and water-propylene glycol and sugaring-out concentration of extracts. *Sep Puri Tech* 2011; **77**(3): 339–346.
- [15] Thakur M, Rana RC, Thakur S. Physicochemical evaluation of *Terminalia chebula* fruits. *J Non Timber Forest Prod* 2008; **15**: 37–42.
- [16] Rangsiwong P, Rangkadilok N, Satayavivad J, Goto M, Shotipruk A. Subcritical water extraction of polyphenolic compounds from *Terminalia chebula* Retz. fruits. *Sep Puri Tech* 2009; **66**: 51–56.
- [17] Muhammad S, Khan BA, Akhtar N, Mahmood T, Rasul A, Hussain I, et al. The morphology, extractions, chemical constituents and uses of *Terminalia chebula*: A review. *J Med Plants Res* 2012; **6**(33): 4772–4775.
- [18] Yoganarasimhan SN. *Medicinal plants of India*. Bangalore: Self Publication; 2000, p. 541.
- [19] Zhang X, Chen C, He S, Ge F. Supercritical-CO₂ fluid extraction of the fatty oil in *Terminalia chebula* and GC-MS analysis. *Zhong Yao Cai* 1997; **20**(9): 463–464.
- [20] Mammen D, Bapat S, Sane R. An investigation to variation in constituents in the fruits of *Terminalia chebula* Retz. at different maturity stages. *Int J Pharm Bio Sci* 2012; **3**(1): 416–419.
- [21] Han Q, Song J, Qiao C, Wong L, Xu H. Preparative isolation of hydrolysable tannins chebulagic acid and chebulinic acid from *Terminalia chebula* by high-speed counter-current

- chromatography. *J Sep Sci* 2006; **29**(11): 1653–1657.
- [22] Bruneton J. *Pharmacognosy, phytochemistry, medicinal plants*. Paris: Lavoisier Publishing; 1995, p.333.
- [23] Khare CP. *Indian herbal remedies: Rational western therapy, Ayurvedic and other traditional usage, Botany*. Berlin: Springer; 2004, p. 451–452.
- [24] Chang CL, Lin CS. Development of antioxidant activity and pattern recognition of *Terminalia chebula* Retzius extracts and its fermented products. *HungKuang J* 2010; **61**: 115–129.
- [25] Naik GH, Priyadarsini KI, Naik DB, Gangabhairathi R, Mohan H. Studies on the aqueous extract of *Terminalia chebula* as a potent antioxidant and a probable radioprotector. *Phytomedicine* 2004; **11**(6): 530–538.
- [26] Mahesh R, Bhuvana S, Begum VM. Effect of *Terminalia chebula* aqueous extract on oxidative stress and antioxidant status in the liver and kidney of young and aged rats. *Cell Biochem Funct* 2009; **27**(6): 358–363.
- [27] Hazra B, Sarkar R, Biswas S, Mandal N. Comparative study of the antioxidant and reactive oxygen species scavenging properties in the extracts of the fruits of *Terminalia chebula*, *Terminalia bellerica* and *Emblica officinalis*. *BMC Comp Alter Med* 2010; **10**: 20.
- [28] Lee HS, Won NH, Kim KH, Lee H, Jun W, Lee KW. Antioxidant effects of aqueous extract of *Terminalia chebula* in vitro and in vitro. *Biol Pharm Bull* 2005; **28**(9): 1639–1644.
- [29] Lee HS, Jung SH, Yun BS, Lee KW. Isolation of chebulic acid from *Terminalia chebula* Retz. and its antioxidant effect in isolated rat hepatocytes. *Arch Toxicol* 2007; **81**(3): 211–218.
- [30] Chen X, Sun F, Ma L, Wang J, Qin H, Du G. In vitro evaluation on the antioxidant capacity of triethylchebulate, an aglycone from *Terminalia chebula* Retz fruit. *Indian J Pharmacol* 2011; **43**(3): 320–323.
- [31] Saleem M, Hushum P, Harkonen K, Pihlaja. Inhibition of cancer cell growth by crude extract and phenolics of *Terminalia chebula* fruit. *J Ethnopharmacol* 2002; **81**: 327–336.
- [32] Reddy DB, Reddy TC, Jyotsna G, Sharan S, Priya N, Lakshmipathi V, et al. Chebulagic acid, a COX–LOX dual inhibitor isolated from the fruits of *Terminalia chebula* Retz., induces apoptosis in COLO–205 cell line. *J Ethnopharmacol* 2009; **124**(3): 506–512.
- [33] Grover IS, Bala S. Antimutagenic activity of *Terminalia chebula* (myroblan) in *Salmonella typhimurium*. *Indian J Exp Biol* 1992; **30**(4): 339–341.
- [34] Gandhi NM, Nayar CKK. Radiation protection by *Terminalia chebula* some mechanistic aspects. *Mol Cell Biochem* 2005; **277**(1–2): 43–48.
- [35] Prasad L, Husain Khan T, Jahengir T, Sultana S. Chemomodulatory effect of *Terminalia chebula* against nickel chloride–induced oxidative stress and tumor promotion response in male Wistar rats. *J Trace Elem Med Biol* 2006; **20**(4): 233–239.
- [36] Lee HS, Jung SH, Yun BS, Lee KW. Isolation of chebulic acid from *Terminalia chebula* Retz. and its antioxidant effect in isolated rat hepatocytes. *Arch Toxicol* 2007; **31**(3): 211–218.
- [37] Tasduq SS, Singh AK, Salti NK, Gupta DK, Suri K. *Terminalia chebula* fruits prevent liver toxicity caused by sub–chronic administration of rifampicin, isoniazid and pyrazinamide (PZA) in combination. *Hum Exp Toxicol* 2006; **25**(3): 11–18.
- [38] Tasaduq SA, Singh K, Sethi S, Sharma SC, Bedi KL, Singh J, et al. Hepatocurative and antioxidant profile of HP–1, a polyherbal phytomedicine. *Hum Exp Toxicol* 2003; **22**(12): 639–645.
- [39] Suchalatha S, Shyamadevi CS. Protective effect of *Terminalia chebula* against experimental myocardial injury induced by isoproterenol. *Indian J Exp Biol* 2004; **42**(2): 174–178.
- [40] Reddy VRC. Cardioprotective activity of the fruit of *Terminalia chebula*. *Fitoterapia* 1990; **61**: 517–525.
- [41] Chang CL, Lin CS, Lai GH, Chen YH, Tuan WC, Hsu CM. Influence of *Terminalia chebula* extracts on the effect of PC12 cell growth. *J Trad Med* 2010; **21**(1): 23–30.
- [42] Minkyun NA, Wan BAE, Kang SS, Min BS, Yoo JK, Yuk OK, et al. Cytoprotective effect on oxidative stress and inhibitory effect on cellular aging of *Terminalia chebula* fruit. *Phytother Res* 2004; **18**: 737–741.
- [43] Lee HS, Koo YC, Suh HJ, Kim KY, Lee KW. Preventive effects of chebulic acid isolated from *Terminalia chebula* on advanced glycation endproduct–induced endothelial cell dysfunction. *J Ethnopharmacol* 2010; **131**(3): 567–574.
- [44] Na M, Bae M, Keng SS, Min BS, Yoo JK, Kamiryo Y, et al. Cytoprotective effect on oxidative stress and inhibitory effect on cellular aging of *Terminalia chebula* fruit. *Phytother Res* 2004; **18**(9): 737–741.
- [45] Kannan VR, Rajasekar GS, Rajesh P, Balasubramanian V, Ramesh N, Solomon EK, et al. Anti–diabetic activity on ethanolic extracts of fruits of *Terminalia chebula* Retz. Alloxan induced diabetic rats. *Am J Drug Discov Dev* 2012; **2**: 135–142.
- [46] Senthilkumar GP, Subramanian SP. Biochemical studies on the effect of *Terminalia chebula* on the levels of glycoproteins in streptozotocin–induced experimental diabetes in rats. *J Appl Biomed* 2008; **6**: 105–115.
- [47] Khan KH, Jain SK. Regular intake of *Terminalia chebula* can reduce the risk of getting typhoid fever. *Adv Biotech* 2009; **8**(9): 10–15.
- [48] Khan KH. The effect of regular intake of *Terminalia chebula* on oxidative stress in mice originated from *Salmonella typhimurium*. *EurAsia J BioSci* 2009; **3**: 113–121.
- [49] Malckzadeh F, Ehsanifar H, Shahamat N, Levin M, Colwell RR. Antibacterial activity of black myrobalan (*Terminalia chebula* Retz.) against *Helicobacter pylori*. *Int J Antimicrob Agent* 2001; **18**(1): 85–88.
- [50] Kim HG, Cho JH, Jeong EY, Lim JH, Lee SH, Lee HS. Growth inhibitory activity of active component from *Terminalia chebula* fruits against intestinal bacteria. *J Food Prot* 2006; **69**(9): 2205–2209.
- [51] Sato Y, Oketani H, Singyouchi K, Ohtsubo T, Kihara M, Shibata H, et al. Extraction and purification of effective antimicrobial constituents of *Terminalia chebula* Retz. against methicillin–resistant *Staphylococcus aureus*. *Bull Pharm Bull* 1997; **20**(4): 401–404.
- [52] Bonjar GH. Antibacterial screening of plants used in Iranian folkloric medicine. *Fitoterapia* 2004; **75**(2): 231–235.
- [53] Aneja KR, Joshi R. Evaluation of antimicrobial properties of fruit extracts of *Terminalia chebula* against dental caries pathogens. *Jundishapur J Microbiol* 2009; **2**(3): 105–111.
- [54] Kannan P, Ramadevi SR, Hopper W. Antibacterial activity of *Terminalia chebula* fruit extract. *Afr J Microbiol Res* 2009; **3**(4): 180–184.
- [55] Rani P, Khullar N. Antimicrobial evaluation of some medicinal plants for their antienteric potential against multi–drug resistant *Salmonella typhi*. *Phytother Res* 2004; **18**(8): 670–673.
- [56] Agrawal A, Gupta A, Choudhary NK, Wadhwa S, Dav K, Goyal S, et al. Antibacterial activity of hydroalcoholic extract of *Terminalia chebula* Retz. on different Gram–positive and Gram–negative Bacteria. *Int J Pharm Biol Arch* 2010; **1**(4): 485–488.
- [57] Bag A, Bhattacharyya SK, Bharati P, Pal NK, Chattopadhyay RR. Antibacterial activity of *Chebulic myrobalan* (fruit of *Terminalia chebula* Retz.) extracts against methicillin resistant *Staphylococcus aureus* and trimethoprim–sulphamethoxazole resistant uropathogenic *Escherichia coli*. *Afr J Plant Sci* 2009; **3**(2): 25–29.
- [58] Bag A, Bhattacharyya SK, Pal NK, Chattopadhyay RR. Synergistic effect of *Terminalia chebula* against multidrug–resistant uropathogenic *Escherichia coli*. *Med Aromatic Plant Sci Biotech*

- 2011; **5**(1): 70–73.
- [59] Barazani VO, Sathiyamoorthy P, Shalev R, Vardy D, Golan GA. Screening of South-Indian medicinal plants for anti-fungal activity. *Phyther Res* 2003; **17**(9): 1123–1125.
- [60] Dutta BK, Rahman I, Das TK. Antifungal activity of Indian plant extracts. *Mycoses* 1998; **41**(11–12): 535–536.
- [61] Mehmood Z, Ahmad I, Mohammad F, Ahmad S. Indian medicinal plants: A potential source of anticandidal Drugs. *Pharm Biol* 1999; **37**(3): 237–242.
- [62] Bonjar GH. Inhibition of Clotrimazole-resistant *Candida albicans* by plants used in Iranian folkloric medicine. *Fitoterapia* 2004; **75**(1): 74–76.
- [63] Jeong AHN, Kim CY, Lee JS, Kim TG, Kim SH, Lee CK, et al. Inhibition of HIV-1 integrase by galloyl glucoses from *Terminalia chebula* and flavonol glycoside gallates from *Euphorbia pekinensis*. *Plant Med* 2002; **68**: 457–459.
- [64] Lee D, Boo K, Woo J, Duan F, Lee K, Kwon T, et al. Anti-bacterial and Anti-viral activities of extracts from *Terminalia chebula* barks. *J Korean Soc Appl Biol Chem* 2011; **54**(2): 295–298.
- [65] Badmaev V, Nowakowski M. Protection of epithelial cells against influenza A virus by plant derived biological response modifier Ledretan-96. *Phytother Res* 2000; **44**(4): 245–249.
- [66] Gambari R, Lampronti L. Inhibition of immunodeficiency type-1 virus (HIV-1) life cycle by medicinal plant extracts and plant derived compounds. *Adv Phytomed* 2006; **2**: 299–311.
- [67] Kurowa M, Nagasaka K, Hirabayashi T, Uyama S, Sato H, Kagiya T, et al. Efficacy of traditional herbal medicines in combination with acyclovir against Herpes Simplex Virus-1 infection *in vitro* and *in vivo*. *Antiviral Res* 1995; **27**(1–2): 19–37.
- [68] Yukawa TA, Kurokawa M, Sato S, Yoshida Y, Kageyama S, Hasegawa T, et al. Prophylactic treatment of cytomegalovirus infection with traditional herbs. *Antiviral Res* 1996; **32**(2): 63–70.
- [69] Vermani K, Garg S. Herbal medicines for sexually transmitted diseases and AIDS. *J Ethnopharmacol* 2002; **80**: 49–66.
- [70] Ma H, Zhao YD, Li K, Kang T. A new alternative to treat swine influenza A virus infection: Extracts from *Terminalia chebula* Retz. *Afr J Microbiol Res* 2010; **4**(6): 497–499.
- [71] Dwivedi S, Dwivedi A, Kapadia R, Kaul S. Anthelmintic activity of alcoholic and aqueous extract of fruits of *Terminalia chebula* Retz. *Ethnobot Leaflets* 2008; **12**: 741–743.
- [72] Sohni YR, Kaimal P, Bhatt RM. The antiamebic effect of crude drug formulation of herbal extracts against *Entamoeba histolytica* *in vitro* and *in vivo*. *J Ethnopharmacol* 1995; **45**(1): 43–52.
- [73] Bagavan A, Rahuman AI, Kamaraj C, Kaushik NK, Mohanakrishnan D, Sahal D. Antiplasmodial activity of botanical extracts against *Plasmodium falciparum*. *Parasitol Res* 2011; **108**(5): 1099–1109.
- [74] Moeslinger T, Friedl R, Volf I, Brunner M, Koller E, Spieckermann PG. Inhibition of inducible nitric oxide synthesis by the herbal preparation Padma 28 in macrophage cell line. *Can J Physiol Pharmacol* 2000; **78**(11): 861–866.
- [75] Nair V, Singh S, Gupta YK. Anti-arthritis and disease modifying activity of *Terminalia chebula* Retz. in experimental models. *J Pharm Pharmacol* 2010; **62**(12): 1801–1806.
- [76] Pratibha N, Saxena VS, Amit A, D'Souza P, Bagchi M, Bagchi D. Anti-inflammatory activities of Aller-7, a novel polyherbal formulation for allergic rhinitis. *Int J Tissue React* 2004; **26**(1–2): 43–51.
- [77] Rege NN, Thatte UM, Dahanukar SA. Adaptogenic properties of six Rasayana herbs used in Ayurvedic medicines. *Phytother Res* 1999; **13**: 275–291.
- [78] Shin TY, Jeong HG, Kim DK, Kim SH, Lee JK, Chae BS, et al. Inhibitory action of water soluble fraction of *Terminalia chebula* on systematic and local anaphylaxis. *J Ethnopharmacol* 2001; **74**: 133–140.
- [79] Maruthappan V, Shree KS. Hypolipidemic activity of Haritaki (*Terminalia chebula*) in atherogenic diet induced hyperlipidemic rats. *J Adv Pharm Tech Res* 2010; **1**: 229–235.
- [80] Israni DA, Patel KV, Gandhi TR. Anti-hyperlipidemic activity of aqueous extract of *Terminalia chebula* and Gaumutra in high cholesterol diet fed rats. *Int J Pharm Sci* 2010; **1**(1): 48–59.
- [81] Tamhane MD, Thorate SP, Rege NN, Dahanukar SA. Effect of oral administration of *Terminalia chebula* on gastric emptying : An experimental study. *J Postgrad Med* 1997; **43** (1): 12–13.
- [82] Sharma P, Prakash T, Kotresha D, Ansari MA, Sahrm UR, Kumar B, et al. Antiulcerogenic activity of *Terminalia chebula* fruit in experimentally induced ulcer in rats. *Pharm Biol* 2011; **49**(3): 262–268.
- [83] Seyyed AM, Ali V, Mohammad KGN, Peyman M. Spasmogenic activity of the seed of *Terminalia chebula* Retz in rat small intestine: *In vitro* and *in vitro* studies. *Malays J Med Sci* 2011; **18**(3): 18–26.
- [84] Carounanidy U, Satyanarayanan R, Velmurugan A. Use of an aqueous extract of *Terminalia chebula* as an anticaries agent: a clinical study. *Indian J Dent Res* 2007; **18**(4): 152–156.
- [85] Li K, Diao Y, Zhang H, Wang S, Zhang Z, Yu B, et al. Tannin extracts from immature fruits of *Terminalia chebula* Fructus Retz. promote cutaneous wound healing in rats. *BMC Comp Alter Med* 2011; **11**: 1–9.
- [86] Vani T, Rajani M, Sarkar S, Shishoo CJ. Antioxidant properties of ayurvedic formulation triphala and its constituents. *Int J Pharmacog* 1997; **35**: 313–317.
- [87] Aher VD. Immunomodulatory effect of alcoholic extract of *Terminalia chebula* ripe fruits. *J Pharm Sci Res* 2010; **2**(9): 539–544.
- [88] Mukherjee P, Roy S, Bhattacharyya S, Debnath PK, Jan U, Pandit S, et al. Clinical study of Triphala- a well known phytomedicine from India. *Indian J Pharmacol Ther* 2006; **5**: 51–54.
- [89] Singh RH, Sinha BN. Clinical and physiological studies on the effect of an indigenous compound Rasayana drug in apparently normal aged person. *J Res Indian Med Yoga Homoeo* 1978; **13**: 1–8.
- [90] Amit VS, Saxena N, Prativa M, Bagchi D, Bagchi J. Safety of novel botanical extract formula for ameliorating allergic rhinitis. *Toxicol Mechanisms Methods* 2004; **13**(4): 253–261.
- [91] Panunto W, Jaijoy K, Lerdvuthisophon N, Lertprasertsuke N, Jiruntanat N, Soonthorncharenon N, et al. Acute and chronic toxicity studies of the water extract from dried fruits of *Terminalia chebula* Retz. in rats. *Int J Appl Res Nat Prod* 2011; **3**(4): 36–43.
- [92] Ji-hoon K, Yun-chang K, Chung-Oui H, Sung-Yong Y, Woojin J, Lee K. Mutagenicity and oral toxicity studies of *Terminalia chebula*. *Phytother Res* 2012; **26**: 39–47.
- [93] Ponnusankar S, Pandit S, Venkatesh M, Bandyopadhyay A, Mukherjee PK. Cytochrome P450 inhibition assay for standardized extract of *Terminalia chebula* Retz. *Phytother Res* 2011; **25**(1): 151–154.
- [94] Arora S, Brits E, Kaur S, Kaur K, Sohi RS, Kumar S, et al. Evaluation of genotoxicity of medicinal plant extracts by the Comet and VITOTOX tests. *J Environ Pathol Toxicol Oncol* 2005; **24**(3): 193–200.
- [95] Rathore HS, Makwana M. Prevention of lead toxicity in *Allium cepa* root tip cells with myrobalan (fruit of *Terminalia chebula* Retz.). *Biochem Cell Arch* 2005; **5**(2): 169–176.
- [96] Rathore HS, Shazia B, Sharma A, Makwana M. Prevention of aluminium chloride- induced mitodepression with myrobalan (fruit of *Terminalia chebula* Retz, Combretaceae) in *Allium cepa* model. *Ethnobot leaflets* 2006; **10**: 272–279.